

ORIGINAL RESEARCH PAPER

Removal of total petroleum hydrocarbons from polluted urban soils of the outskirts of Ahvaz, southwestern Iran

A. Takdastan¹, M. Kardani^{2,}, H. Janadeleh³*

¹ Associate Professor, Department of Environmental Health, Environmental Technologies Research Center, Jundishapur University of Medical Sciences, Ahvaz, Iran

² Department of Environmental Science, Khuzestan Science and Research Branch, Islamic Azad University, Ahvaz, Iran

³ Department of Environmental Science, Ahvaz Branch, Islamic Azad University, Ahvaz, Iran

Receive Date 23 December 2016; Revise Date 8 February 2017; Accept Date 24 March 2017; Publish Date 1 April 2017

ABSTRACT: Earlier phases of economic expansion and urban development have resulted in significant sources of urban soil contamination. Petroleum hydrocarbons are one of the most common groups of persistent organic contaminants in the environment. In this study, two types of treatment in 3 concentrations were prepared that were included plant treated by 1% oil pollution, treatment by 1% contamination without plant (as control), plant treated by 5% oil pollution, the 5% pollution treatment without plant (control), 10% oil pollution treatment with plant and 10% treatment without plant (control) that 3 replicates were prepared for each treatment. The obtained extracts were concentrated to 1 mL under a gentle stream of nitrogen gas, and then 2 µg of the sample was injected into a UNICAM 610 series gas chromatograph equipped with a flame ionization detector. Primary Total petroleum hydrocarbons amount in 1%, 5% and 10% concentration was respectively: 9027.40 mg/kg, 49599.03 mg/kg and 99548.28 mg/kg. After 4 months its amount in different concentration with plant was 126.43 mg/kg, 4463.92 mg/kg and 19611.50 mg/kg. The best total petroleum hydrocarbons removal efficiency was observed in all concentration at 120th day. The results of this study showed that *vetiver* can remove petroleum hydrocarbons from contaminated soils effectively.

KEYWORDS: Ahvaz city; Flame ionization detector (FID); Petroleum hydrocarbons; Removal; Soil; Total petroleum hydrocarbons (TPH)

INTRODUCTION

The urban environmental quality is of basic importance because the majority of people now live in cities. Urban soils are therefore an important indicator of human exposure to pollution such as total petroleum hydrocarbons and heavy metal in the urban terrestrial environment (Nriagu and Pacyna, 1988). Urban soils can vary widely over short distances, compared to naturally developed and anthropogenically influenced soils. Most urban soils are relatively young, resulting from soil exchange and mixture due to construction

activities (Norra *et al.*, 2001; Janadeleh and Kardani, 2016). Petroleum hydrocarbons are one of the most common groups of persistent organic contaminants in the environment (Collins, 2007; Cook and Hesterberg, 2013; El-Sheekh and Hamouda, 2014; Janadeleh and Jahangiri, 2016). Petroleum hydrocarbon contamination of soil occurs through extraction, accidents, transportation, leakage from tanks, pipeline ruptures, consumption and refining (Soleimani *et al.*, 2010; Kokyo *et al.*, 2013). Large-scale contamination of soil and sediments with petroleum hydrocarbons is a serious problem of global scale, primarily in countries that

✉ *Corresponding Author Email: m.kordani86@yahoo.com
Tel.: +98 916 6206 651; Fax: +98 61 3320 7329

produce, transport and refine oil (Muratova et al., 2008; Basumatary et al., 2012). Over the past centuries, several factors, such as rapid growth of population, modern agricultural activities, waste disposal, mining, and industrialization, have significantly contributed to extensive soil contamination (Eapen and D'souza, 2005; Singh and Jain, 2003). Soil is the foundation of terrestrial ecosystems and a nonrenewable fundamental agricultural resource, inextricably linked to productivity, land development and environmental quality. Interest in phytoremediation as a method to solve chemical contamination has been growing rapidly in recent years (Clemens et al., 2002; Dhankher et al., 2002; Uera et al., 2007). Phytoremediation is the process of applying plants for removing contaminants from the soil, surface and ground waters, which has been developed during the past decade (Baneshi et al., 2014; Moreira et al., 2013; Patil and Jadhav, 2013). It is believed that the first step towards phytoremediation is to find an appropriate set of plants which enable them to grow on contaminated sites, although optimizing agronomic practice should also be considered. In contrast, phytoremediation is considered to be a less environmentally destructive, more aesthetically pleasing and solar powered treatment system. Besides, it is more cost-effective than most of the alternatives (Cunningham et al., 1996; Brandt et al., 2006; Janadeleh et al., 2017). This method is referred to as plant-assisted remediation or phytoremediation, and it also has the benefit of contributing to site restoration when remedial action is ongoing. *C. zizanioides* (L.) Roberty, commonly known as *vetiver*, is a fast-growing perennial tussock grass member of the family Poaceae; it is considered sterile outside its natural habitat. *Vetiver* is grown worldwide for perfumery, agriculture, and bioengineering where it is used for soil and water conservation (Barbera et al., 2014). *Vetiveria zizanioides* native from Indian subcontinent and has the potential to meet all the criteria required for phytoremediation (Abaga, et al., 2014; Danh et al., 2009). *Vetiver* showed promise in removing various environmental contaminants from both aqueous media and soil (Danh et al., 2009; Datta et al., 2011; 2013; Janadeleh et al., 2016). Many plant species are sensitive to petroleum contaminants (Chaîneau, et al., 1997; Huang et al., 2004). "According to Merkl et al. (2004) a growth rate reduction of beans and wheat by more than 80%. Significant reduction of plant biomass by the presence of petroleum hydrocarbons

has also been reported. Ahwaz is located in the southwest of Iran, and the capital of Khuzestan province. Ahwaz has a desert climate with long, very hot summers and mild, short winters. Ahwaz is consistently one of the hottest cities on the planet during the summer, with summer temperatures regularly at least 45 degrees Celsius, sometimes exceeding 50 degrees Celsius, with many sandstorms and duststorms common during the summer period. However, in winters, the minimum temperature can fall to around +5 degrees Celsius. Winters in Ahwaz have no snow. The average annual rainfall is around 230 mm. This study has been carried out around Ahwaz City of Iran in 2015.

MATERIALS AND METHODS

Method of artificially soil contamination

In present study the Soil samples were collected from oil field around Ahwaz city. Then in order to infect it by 1%, 5% and 10% concentration of soil contamination, soil was flat in a thin layer. Then the crude oil was dissolved in acetone in ratio of 1 to 3 and was added to soil in the form of spray. 30 cm Pot was filled by contaminated soil with various concentrations of contaminant. Soil of each pot was weighed 7Kg and was ready for planting. In this study, two types of treatment in 3 concentrations were prepared that were included plant treated by 1% oil pollution, treatment by 1% contamination without plant (as control), plant treated by 5% oil pollution, the 5% pollution treatment without plant (control), 10% oil pollution treatment with plant and 10% treatment without plant (control) that 3 replicates were prepared for each treatment.

Sampling and sample digestion

This study was evaluated in 5 period times (15, 30, 60, 90, 120 days). Some soil samples from each pot were taken for the analysis, and keep in the zippers bags and then were transferred to a laboratory for analysis. Some physical and chemical properties of the clean soil as well as contaminated soil used in this study are presented in Table 1. Phosphorus was measured by Olsen P extracting solution (0.5 M NaHCO₃, pH 8.5); total nitrogen was measured by Kjeldahl digestion; EC was measured by a conductivity meter in a soil-water extract (1:5 soil: water ratio), and pH was analyzed by a glass electrode using a 1:5 soil: water ratio (Dewis and Freitas, 1970; Testing and Materials, 2000). Total Volatile Solids was measured by Using of Gravimetric according to method 1684 EPA (Telliard, 2001).

Table 1: Physical and chemical characteristics of the clean soil used in the current study

Parameter	Value	Measurement method
pH	7.27	1:5 soil/water slurry
Electrical Conductivity(ds/m)	8.50	1:5 soil/water slurry
Total N	0.087	Kjeldahl
Phosphorus (mg/kg)	3.62	Olsen
Clay	9.00	Hydrometer measurement
Sand	80.20	Hydrometer measurement
Silt	10.80	Hydrometer measurement
Inorganic matter	92.44	Gravimetric
Organic matter	7.60	Gravimetric

TPH analysis

For TPH analysis, soil samples were air dried and stored at 4 °C prior to extraction and analysis. Ultrasonic extraction was performed using dichloromethane solvent. Dichloromethane (10 mg) was added to about 5 g of contaminated soil and then it was placed in an ultrasonic water bath for 3 min at room temperature (Walters *et al.*, 1989). The obtained extracts were concentrated to 1 mL under a gentle stream of nitrogen gas, and then 2 µg of the sample was injected into a UNICAM 610 series gas chromatograph equipped with a flame ionization detector (FID). The column used for analysis was DB-5 with 30 m length, 0.25 mm internal diameter, and 0.2 µm thickness of film. The injector and FID detector temperatures were adjusted at 280 °C and 340°C, respectively. Initial column temperature was adjusted at 50 °C for 5 min, and then increased to 250°C with 10 °C min⁻¹ slope and remained at 250 °C for 40 min.

HPC analysis

The heterotrophic plate count (HPC), formerly known as the standard plate count, is a procedure for estimating the number of live heterotrophic bacteria in water. This test can provide useful information about water quality and supporting data on the significance of coliform test results. High concentrations of the general bacterial population may hinder the recovery of coliforms.

The HPC is useful in judging the efficiency of various treatment processes for both drinking water and swimming pools, and for checking the quality of finished water in a distribution system. 8241 Pour Plate, Heterotrophic Bacteria Plate Count Agar, Count CFU/MI (HACH Company, 2012)

Statistical analysis

In this study the significance of differences observed among the mean values was tested by one-way ANOVA. Significance level was considered at P = 0.05. All statistical analyses were performed using SPSS 17.

RESULTS AND DISCUSSION

Physical and Chemical analysis of both soil contaminated by petroleum hydrocarbons and untamed some physical and chemical properties of the soil were measured by standard methods (Table 1). According to the results of present study the texture of soil had 50% porosity and sandy loam, which its sand percentage was reported 80.2%. Results of TPH amount in soil without plant and with plant presence is showed in Table 2.

TPH removal rate during phytoremediation process

The results of TPH concentration in soil samples in different time period during the phytoremediation

Phytoremediation of TPH from polluted soil

Table 2: Different concentration of studied contaminated soil (mg/kg)

Contamination percentage made by artificially	Primary Concentration	Concentration in plant presence	TPH removal percentage
1%	9027.40	126.43	98.6
5%	49599.03	4463.92	91
10%	99548.28	19611.50	80.3

Table 3: TPH removal rate in different treatments (mg/kg)

Soil sample	Treatment with plant			Treatment without plant (control)		
	1%	5%	10%	1%	5%	10%
Contamination percentage						
Time interval (day)						
0	9027.40	49599.03	99548.27	9027.40	49599.03	99548.27
15	3520.07	30751.48	71674.81	6138.64	36802.49	91385.32
30	1579.89	25047.50	63213.28	4965.07	35711.31	88299.32
60	875.78	11755.59	40615.73	4694.25	30652.21	71674.76
90	343.12	9542.91	30481.72	3430.42	21575.58	67692.83
120	126.43	4463.92	19611.50	2979.05	19343.63	63013.06

process with *Vetiveria zizanioides* are presented in Fig. 1. The rate of TPH removal was evaluated in control treatment with plant and 1%, 5% and 10% concentration. The Results from this study suggest that the highest TPH removal during 4 months in all treatments with plant had significantly ($P < 0.05$) difference in compared with control treatments. Primary TPH amount in 1%, 5% and 10% concentration was respectively: 9027.40 mg/kg, 49599.03 mg/kg and 99548.28 mg/kg. After 4 months its amount in different concentration with plant was 126.43 mg/kg, 4463.92 mg/kg and 19611.50 mg/kg. In addition, Peng *et al.* (2009) noted 41.61–63.2% removal of TPH by *Mirabilis Jalapa*, during 127 days (Peng *et al.*, 2009). The best TPH removal efficiency was observed in all concentration at 120th day and in treatments with plant. Although plants with highly branched fine fibrous root systems which have higher

total rhizosphere volume have been reported to enhance biodegradation of organic contaminants more than plants with taproot systems (Soleimani *et al.*, 2010; Aprill and Sims, 1990). Euliss *et al.* (2008) found out that *Carex stricta*, *Panicum virgatum* and *Tripsacum dactyloides* significantly reduced TPH by 70% after 1 year of growth (Euliss *et al.*, 2008). Also, the lowest TPH removal rate was observed in all concentration at 15th day and control treatments. In 1%, 5% and 10% of oil pollution TPH removal rate was 32%, 25.8% and 8.2%, respectively. 1% oil pollution treatment had no correlation with time. 5% oil pollution treatment had positive correlation with time at 1% level, 10% oil pollution treatment had negative correlation with time at 1% level. The results showed that TPH removal amount was higher than controls in all treatment with plant in all contamination concentration treatments.

Organic solids changes rate during phytoremediation process

The results of Volatile Solids concentrations at different time period during the phytoremediation process with *Vetiveria zizanioides* were presented in Fig. 2. The results showed that 1% treatment at 30, 60, 90 and 120 days had significant differences ($P < 0.05$) in

compared to control. 5% treatment at 60, 90 and 120 days had significant difference ($P < 0.05$). 10% treatment at 90 and 120 days showed significant differences ($P < 0.05$) with control. The amount of organic solid reduction in 1%, 5% and 10% oil pollution concentration was 62.96%, 65.49% and 66.41%, respectively, and 10% treatment had maximum of organic solid reduction.

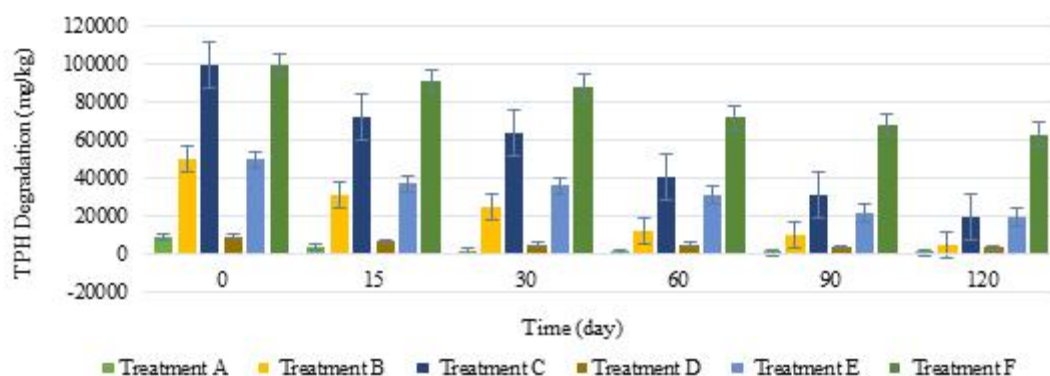


Fig. 1: Degradation of TPH pollutants at different time points during the phytoremediation process with *Vetiveria zizanioides* on 4 months. Values are means + standard deviation (SD). Treatment A: TPH -contaminated soil with concentration 1% (with plant). Treatment B: TPH-contaminated soil with concentration 5% (with plant). Treatment C: TPH-contaminated soil with concentration 10% (with plant). Treatment D: TPH-contaminated soil with concentration 1% (no plant). Treatment E: TPH-contaminated soil with concentration 5% (no plant). Treatment F: TPH-contaminated soil with concentration 10% (no plant).

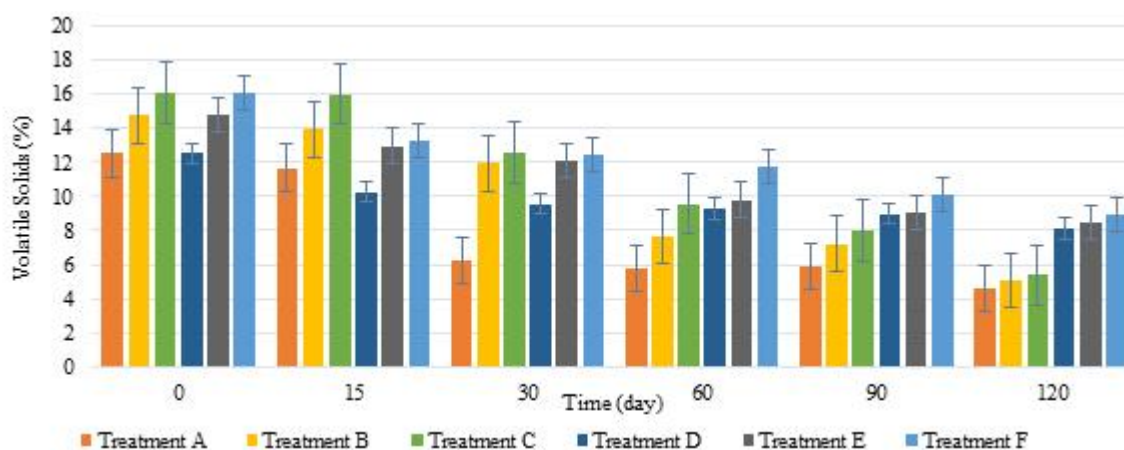


Fig. 2: Concentration of Volatile Solids at different time points during the phytoremediation process with *Vetiveria zizanioides* on 4 months. Values are means + standard deviation (SD). Treatment A: TPH-contaminated soil with concentration 1% (with plant). Treatment B: TPH-contaminated soil with concentration 5% (with plant). Treatment C: TPH-contaminated soil with concentration 10% (with plant). Treatment D: TPH-contaminated soil with concentration 1% (no plant). Treatment E: TPH-contaminated soil with concentration 5% (no plant). Treatment F: TPH-contaminated soil with concentration 10% (no plant).

Changes of microbial activity during phytoremediation process

Bacterial cell in CFU/g of soil (10^6) on Day0 and Day120 were showed in Fig. 3. The soil heterotrophic microbial populations studied were counted at the first day and 120th days. Studies show that all treatments were significantly different to each other at 15th days and 120th days ($P < 0.05$) and only 5% treatment had significantly different with controls ($P < 0.05$). Soil microbial population increased during 120 days in treatments with plants. Number of microbial population in 1%, 5% and 10% contamination treatments with plant was 8.6×10^6 CFU/g, 9.46×10^6 CFU/g and 8.1×10^6 CFU/g, respectively. Organic matter with hydrocarbon in soil as food resource for bacteria was reason of increasing in number of heterotrophic bacterial populations during 120 days. But number of heterotrophic bacteria in 10% oil contamination treatments had reduction in compared to other treatments that was due to more contamination and inhibition of bacteria growth.

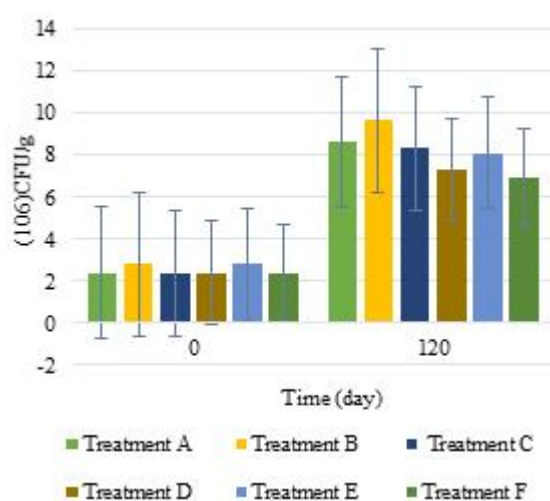


Fig. 3: Bacterial cell in CFU/g of soil (10^6) on Day 0 and Day120 Values are means + standard deviation (SD). Treatment A: TPH-contaminated soil with concentration1% (with plant). Treatment B: TPH-contaminated soil with concentration5% (with plant). Treatment C: TPH-contaminated soil with concentration10% (with plant). Treatment D: TPH-contaminated soil with concentration1% (no plant). Treatment E: TPH-contaminated soil with concentration5% (no plant). Treatment F: TPH-contaminated soil with concentration10% (no plant).

CONCLUSION

The results showed that with *Vetiveria zizanioides* can remove petroleum hydrocarbons from contaminated soils. Due to the characteristics of *Vetiveria zizanioides* plants with high tolerance to extreme climate change, long-term drought and as well as a wide geographic area in the world and also, according to the results of this study the *Vetiveria zizanioides* can be using for removal petroleum hydrocarbons in soil. The use of vegetation as a feasible remediation approach for soils contaminated with petroleum hydrocarbons may become attractive in Iran as well as in other developing countries because it is inexpensive and requires minimum maintenance and little management.

ACKNOWLEDGEMENT

The authors acknowledge the financial support provided by the National Iranian Drilling Company.

CONFLICT OF INTEREST

There is no conflicts of interest for publication of the manuscript.

REFERENCES

- Abaga, N.O.Z.; Dousset, S.; Munier-Lamy, C.; Billet, D., (2014). Effectiveness of Vetiver Grass (*Vetiveria Zizanioides* L. Nash) for Phytoremediation of Endosulfan in Two Cotton Soils from Burkina Faso. International journal of phytoremediation., 16(1): 95-108 (14 pages).
- Andra, S.S.; Datta, R.; Sarkar, D.; Makris, K.C.; Mullens, C.P.; Sahi, S.V.; Bach, S.B., (2010). Synthesis of phytochelatin in vetiver grass upon lead exposure in the presence of phosphorus. Plant Soil, 326(1-2): 171-185 (15 pages).
- Aprill, W.; Sims, R.C., (1990). Evaluation of the use of prairie grasses for stimulating polycyclic aromatic hydrocarbon treatment in soil. Chemosphere, 20(1): 253-265 (13 pages).
- Baneshi, M.M.; Rezaei Kalantary, R.; Jonidi Jafari, A.; Nasser, S.; Jaafarzadeh, N.; Esrafil, A., (2014). Effect of bioaugmentation to enhance phytoremediation for removal of phenanthrene and pyrene from soil with Sorghum and *Onobrychis sativa*. J. Environ. Health Sci. Eng., 12(1): 12-24 (13 pages).
- Barbera, A.C.; Borin, M.; Cirelli, G.L.; Toscano, A.; Maucieri, C., (2014). Comparison of carbon balance in Mediterranean pilot constructed wetlands vegetated with different C4 plant species. Environ. Sci. Pollut. Res., 22(4): 2372-2383 (12 pages).
- Basumatary, B.; Saikia, R.; Bordoloi, S.; Das, H.C.; Sarma, H.P., (2012). Assessment of potential plant species for phytoremediation of hydrocarbon contaminated areas of upper Assam, India. J. Chem. Technol. Biotechnol., 87(9): 1329-1334 (6 pages).
- Brandt, R.; Merkl, N.; Schultze-Kraft, R.; Infante, C.; Broll, G., (2006). Potential of vetiver (*Vetiveria zizanioides* (L.) Nash) for phytoremediation of petroleum hydrocarbon-contaminated soils in Venezuela. Int. J. Phytorem., 8(4): 273-284 (12 pages).
- Chaineau, C.; Morel, J.; Oudot, J., (1997). Phytotoxicity and plant uptake of fuel oil hydrocarbons. J. Environ. Qual., 26(6): 1478-1483 (6 pages).

- Clemens, S.; Palmgren, M.G.; Krämer, U., (2002). A long way ahead: understanding and engineering plant metal accumulation. *Trends Plant Sci.*, 7(7): 309-315 (7 pages).
- Cook, R.L.; Hesterberg, D., (2013). Comparison of trees and grasses for rhizoremediation of petroleum hydrocarbons. *Int. J. Phytorem.*, 15(9): 844-860 (27 pages).
- Cunningham, S.D.; Anderson, T.A.; Schwab, A.P.; Hsu, F.C., (1996). Phytoremediation of soils contaminated with organic pollutants. *Adv. Agron.*, 56(1): 55-114 (60 pages).
- Danh, L.T.; Truong, P.; Mammucari, R.; Tran, T.; Foster, N., (2009). Vetiver grass, *Vetiveria zizanioides*: a choice plant for phytoremediation of heavy metals and organic wastes. *Int. J. Phytorem.*, 11(8): 664-691 (28 pages).
- Datta, R.; Das, P.; Smith, S.; Punamiya, P.; Ramanathan, D.M.; Reddy, R.; Sarkar, D., (2013). Phytoremediation potential of vetiver grass [*Chrysopogon zizanioides* (L.)] for tetracycline. *Int. J. Phytorem.*, 15(4): 343-351 (9 pages).
- Datta, R.; Quispe, M.A.; Sarkar, D., (2011). Greenhouse study on the phytoremediation potential of vetiver grass, *Chrysopogon zizanioides* L., in arsenic-contaminated soils. *Bull. Environ. Con. Tox.*, 86(1): 124-128 (5 pages).
- Dewis, J.; Freitas, F., (1970). Physical and chemical methods of soil and water analysis. *FAO Soils Bulletin.*, 10.
- Dhankher, O.P.; Li, Y.; Rosen, B.P.; Shi, J.; Salt, D.; Senecoff, J.F.; Sashti, N.A.; Meagher, R.B., (2002). Engineering tolerance and hyperaccumulation of arsenic in plants by combining arsenate reductase and α -glutamylcysteine synthetase expression. *Nat. Biotechnol.*, 20(11): 1140-1145 (6 pages).
- Eapen, S.; D'souza, S., (2005). Prospects of genetic engineering of plants for phytoremediation of toxic metals. *Biotechnol. Adv.*, 23(2): 97-114 (18 pages).
- El-Sheekh, M.; Hamouda, R., (2014). Biodegradation of crude oil by some cyanobacteria under heterotrophic conditions. *Desalin. Water Treat.*, 52(7-9): 1448-1454 (7 pages).
- Euliss, K.; Ho, C.-h.; Schwab, A.; Rock, S.; Banks, M.K., (2008). Greenhouse and field assessment of phytoremediation for petroleum contaminants in a riparian zone. *Bioresour. Technol.*, 99(6): 1961-1971 (11 pages).
- HACH Company, (2012). Heterotrophic Bacteria, Pour Plate, Plate Count Agar, Method 8241. *The Hach Water Analysis Handbook.*, USA. (8 pages).
- Huang, X.-D.; El-Alawi, Y.; Penrose, D.M.; Glick, B.R.; Greenberg, B.M., (2004). A multi-process phytoremediation system for removal of polycyclic aromatic hydrocarbons from contaminated soils. *Environ. Pollut.*, 130(3): 465-476 (11 pages).
- Janadeleh, H.; Kardani, M., (2016). Heavy Metals Concentrations and Human Health Risk Assessment for Three Common Species of Fish from Karkheh River, Iran. *Iranian J. Toxicol.*, 10(6): 31-37 (7 pages).
- Janadeleh, H.; Hosseini Alhashemi, A.; S. Nabavi., (2016). Investigation on concentration of elements in wetland sediments and aquatic plants. *Global J. Environ. Sci. Manage.*, 2(1): 87-93 (7 pages).
- Janadeleh, H.; Jahangiri, S., (2016). Study of Contamination and Risk Assessment of Heavy Metal in Fish (*Otolithes ruber*) and Sediments from Persian Gulf. *J. Community Health Res.*, 5(3): 159-181 (23 pages).
- Janadeleh, H.; Kameli, M.A.; Boazar, C., (2017). Seasonal variations of metal pollution and distribution, sources, and ecological risk of polycyclic aromatic hydrocarbons (PAHs) in sediment of the Al Hawizah wetland, Iran. *Hum. Ecol. Risk Assess.: An Int. J.*, 1-18 (19 pages).
- Kokyo, O.; Tao, L.; Hongyan, C.; Xuefeng, H.; Chiquan, H.; Lijun, Y.; Yonemochi, S., (2013). Development of profitable phytoremediation of contaminated soils with biofuel crops. *Journal of Environmental Protection.* 4(4A): 58-64 (7 pages).
- Merkel, N.; Schultze-Kraft, R.; Infante, C., (2004). Phytoremediation in the tropics—the effect of crude oil on the growth of tropical plants. *Bioresour. J.*, 8(3-4): 177-184 (8 pages).
- Moreira, I.T.; Oliveira, O.M.; Triguís, J.A.; Queiroz, A.F.; Ferreira, S.L.; Martins, C.M.; Silva, A.C.; Falcão, B.A., (2013). Phytoremediation in mangrove sediments impacted by persistent total petroleum hydrocarbons (TPH's) using *Avicennia schaueriana*. *Mar. Pollut. Bull.*, 67(1): 130-136 (7 pages).
- Muratova, A.Y.; Dmitrieva, T.; Panchenko, L.; Turkovskaya, O., (2008). Phytoremediation of Oil-Sludge-Contaminated Soil. *Int. J. Phytorem.*, 10(6): 486-502 (17 pages).
- Nriagu, J.O.; Pacyna, J.M., (1988). Quantitative assessment of worldwide contamination of air, water and soils by trace metals. *Nat.*, 333: 134-139 (4 pages).
- Norra, S.; Weber, A.; Kramar, U.; Stiben, D., (2001). Mapping of trace metals in urban soils: the example of Muhlburg/Karlsruhe, Germany. *J. Soils Sediments.*, 1(2): 77-97 (21 pages).
- Patil, A.V.; Jadhav, J.P., (2013). Evaluation of phytoremediation potential of *Tagetes patula* L. for the degradation of textile dye reactive blue 160 and assessment of the toxicity of degraded metabolites by cytogenotoxicity. *Chemosphere*, 92(2): 225-232 (8 pages).
- Peng, S.; Zhou, Q.; Cai, Z.; Zhang, Z., (2009). Phytoremediation of petroleum contaminated soils by *Mirabilis jalapa* L. in a greenhouse plot experiment. *J. Hazard. Mater.*, 168(2): 1490-1496 (7 pages).
- Singh, O.; Jain, R., (2003). Phytoremediation of toxic aromatic pollutants from soil. *Appl. Microbiol. Biotechnol.*, 63(2): 128-135 (8 pages).
- Soleimani, M.; Afyuni, M.; Hajabbasi, M.A.; Nourbakhsh, F.; Sabzalian, M.R.; Christensen, J.H., (2010). Phytoremediation of an aged petroleum contaminated soil using endophyte infected and non-infected grasses. *Chemosphere*, 81(9): 1084-1090 (7 pages).
- Telliard, W., (2001). Method 1684: Total, fixed, and volatile solids in water, solids, and biosolids. *US Environmental Protection Agency.* Washington.

Phytoremediation of TPH from polluted soil

Uera, R.B.; Paz-Alberto, A.M.; Sigua, G.C., (2007). Phytoremediation potentials of selected tropical plants for ethidium bromide. *Environ. Sci. Pollut. Res. Int.*, 14(7): 505-509 (**5 pages**).

Walters, G.; Zilis, K.; Wessling, E.; Hoffman, M., (1989). Analytical methods for petroleum hydrocarbons. *Volatile Organics Control*, 620-623 (**4 pages**).